

**SAFETY MANUAL
AND
STANDARD OPERATING PROCEDURES**

**FOR THE
OCS Cytometry Core**

**NEW YORK UNIVERSITY
SCHOOL OF MEDICINE**

Skirball Institute, Laboratory 3-9 back

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Approval Date:		Effective Date:	
Approved by:			
Peter Lopez			
Director			
Gerry Griffin			
Associate Director, Environmental Health and Safety			
Mark Olmsted			
Environmental Specialist II, Environmental Health and Safety			
Written by:			
Michael Gregory, Laboratory Manager, and Keith Kobylarz, Sr. Lab Tech			
Mark Olmsted			
Environmental Specialist II, Environmental Health and Safety			

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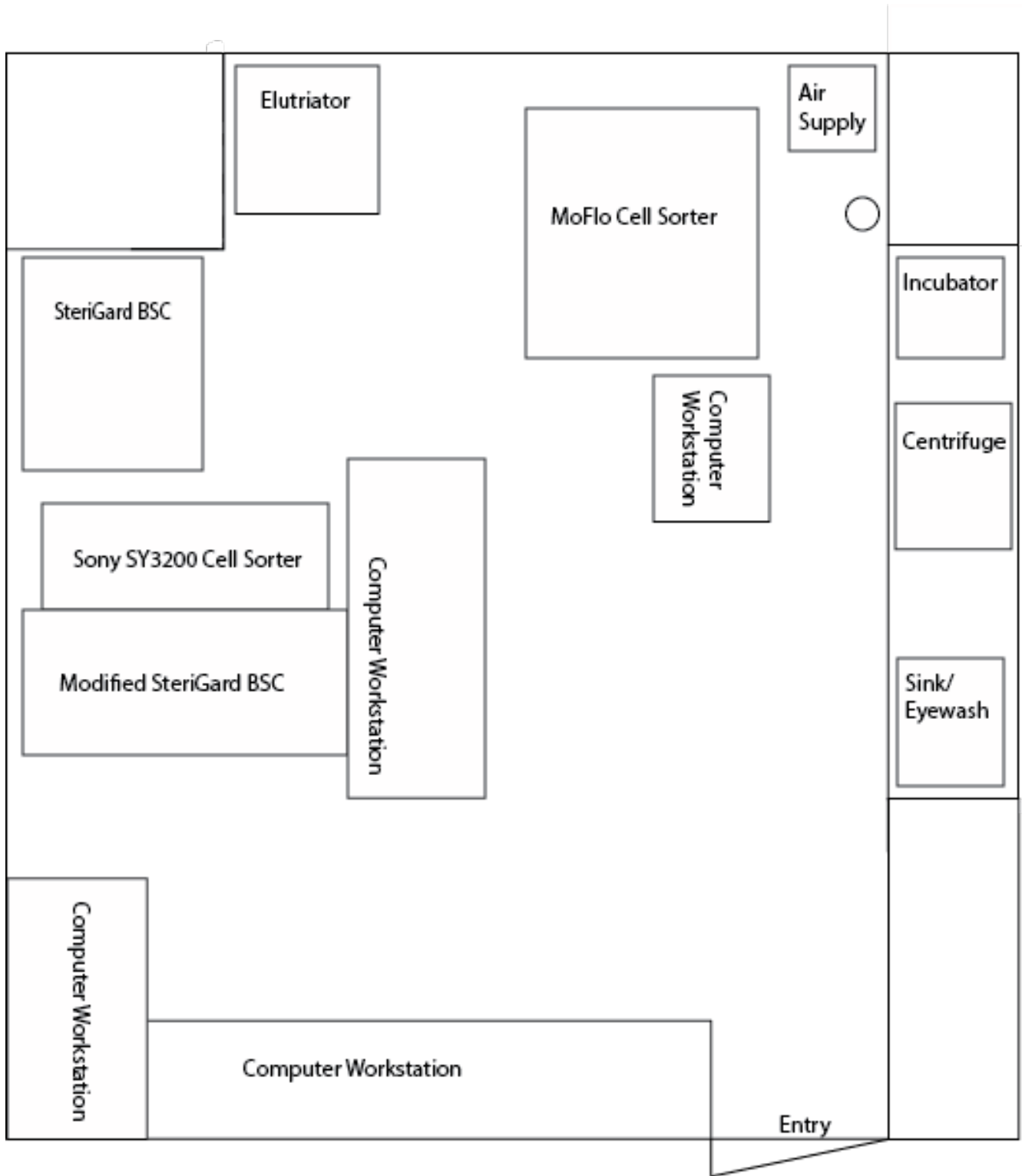
List of Key Personnel

Name	Title	Extension	Mobile
Peter Lopez	Director	x30635	646-357-0168
Michael Gregory	Technical Director	x35907	516-641-5185
Kamilah Ryan	Asst Research Scientist	x35907	917-715-5337
Yulia Chupalova	Asst Research Scientist	x35907	203-570-3649
Keith Kobylarz	Sr. Lab Technician	x35907	718-664-8633

Important Telephone Numbers

Note: <i>In an emergency, Communications can page personnel from key departments</i>	
Any Medical Center Emergency	x33911
Building Services	x35071
Communications	x37403
NYULMC Emergency Department (ED)	x35550
Occupational Health Services	x35020
Environmental Health and Safety (EH&S)	x35159
Mark Olmsted, Environmental Specialist II	x35161
Gerry Griffin, Associate Director	x36944
NYULMC Facilities Management	x35275
Poison Control	(212) 764-7667
Radiation Safety	x36888
Security	x73000
Environmental Services	X34930
Institutional Biosafety Committee (IBC)	(646) 754-5258

Skirball BSL-2 Laboratory Layout



1. Background

1.1. General Information

The BSL-2 cell-sorting laboratory is located on the 3rd floor of the Skirball Institute, in the back room of laboratory 3-9. A locking door restricts access to this facility. The facility includes a front room, used to store laboratory supplies and housing instruments not requiring containment. The main laboratory room contains three Class II re-circulating biosafety cabinets, one housing the cell sorting instrument and one for sample preparation/tissue culture. Special instrumentation is employed in this area to contain potential aerosols and reduce operator exposure. For more detailed information please see the CDC web site: <http://www.cdc.gov/od/ohs/biosfty/biosfty.htm>

2. Risk to Lab Personnel

2.1. Definition of Biohazardous Specimens

All unfixed human and primate cell suspensions and tissues must be treated as potentially infectious, and handled in accordance with universal precautions for blood borne pathogens (i.e., handle as if infected with HIV, HBV, HCV etc.). This applies to cultured cell lines as well as primary tissue suspensions (e.g., blood, bone marrow, cells derived from solid organs). It also applies to nonhuman cells that have been deliberately infected with known or potential human pathogens. Although standard BSL-2 working conditions are usually acceptable for handling such specimens, the potential of cell sorters to generate high levels of aerosolized microdroplets require additional precautions. For the purposes of high speed cell sorting, specimens considered to be potentially biohazardous include all of the following:

- Suspensions of primary human or primate cells from blood or other tissues.
- Cultured and in vitro passaged human or primate cell lines. Note that with few if any exceptions, established human cell lines may fall into the “potentially biohazardous” category, and therefore cannot be sorted unless specific recommendations for sorting biohazardous specimens are followed.
- Primary cells or cell lines that have been transformed with an immortalization agent that has the potential to transform human cells, such as Epstein-Barr virus or a potentially oncogenic retrovirus or lentivirus.
- Any samples known to contain or have been exposed to infectious pathogens normally handled at BSL-2 conditions. This includes agents such as viruses (HIV, HCV, HBV, CMV, EBV, influenza, etc.), bacteria (*Listeria*, BCG and other attenuated mycobacteria, staphylococci, streptococci, various Gram negative pathogens, etc.), fungi (*Cryptococcus*, *histoplasma*, *aspergillus*) and protozoa (*Toxoplasma*, some plasmodia, *cryptosporidia*, etc.).

2.2. High-speed Cell Sorting

High speed droplet based cell sorters can generate large amounts of aerosols, and recently published standards now specify a much higher level of biocontainment for cell sorting of unfixed human cells or other potentially biohazardous samples than have been traditionally followed. “If aerosol containment is incomplete, the safety features of the cell sorter must be modified such that no escape of aerosol can be detected. Alternately, sorters can be placed

inside a biosafety containment cabinet” (Ref: I Schmid et al., International Society for Analytical Cytology Biosafety Standard for Sorting of Unfixed Cells. Cytometry Part A, 71A:414-437 (2007)).

Sorting of samples that represent potential toxic or infectious exposures via the aerosol route therefore require special procedures and laboratory conditions. This is true even for agents that are normally handled under standard BSL-2 laboratory conditions, such as primary human cell suspensions or cell lines. The heightened concern in the case of cell sorting arises from the possibility that cells or microorganisms may be delivered directly into the lungs of personnel in the vicinity of a cell sorter. In theory, this could increase the risk of infection with an occult pathogen, transfer of genetic material, sensitization to antigens or other potentially harmful effects. Although such adverse effects have not been documented as a consequence of exposure to aerosols during cell sorting, there is sufficient concern about this to warrant the implementation of procedures to eliminate any excess risk to personnel.

3. Containment

3.1. Facility Layout

3.1.1. The Skirball OCS Cytometry Core Laboratory is a Biosafety Level 2 (BSL 2) certified facility that is located in NYUMC’s Skirball Institute, Laboratory 3-9.

3.1.2. The facility is cleaned and maintained by the laboratory staff supplemented by Collins Building Services (CBS) a contractor managed by Environmental Services.

3.2. Laboratory Facilities

3.2.1. The laboratory has one sink for hand washing near the entrance to the back room of Skirball 3-9. An eye wash station is located near the sink. An emergency shower is at main the entrance to Lab 3-9. The eyewash within the lab can also serve the function of the emergency shower.

3.2.2. The eye wash station and emergency shower are maintained and inspected by the NYULMC Facilities Department (212-263-4930).

3.3. Biosafety Cabinets (BSCs) and Aerosol Management Unit

3.3.1. There is one three-foot Baker SteriGardIII Advance Class II BSC, one modified Baker SteriGard Class II BSC, and one Walker Class II BSC located in the facility. The modified Baker SteriGard cabinet houses the Sony SY3200 cell sorter and provides extra bench space for sample preparation. The Walker Class II BSC houses the Beckman Coulter MoFlo XDP cell sorter. The Baker Sterigard III Advance Class II BSC is used for cell culture, centrifuge elutriation sample preparation, and flow cytometry sample preparation.

Name/ Room#	Room	Model	BSC Serial No.
Baker SteriGard III Advance	SK 3-9	SG303	91620
Baker Modified SteriGard	SK 3-9	SG405A-HE	107613
Walker Medical Safety Cabinet	SK 3-9	Class II MSC Walker	WSC 1751A

3.3.2. Certification of BSCs - Environmental Health and Safety (EH&S) retains a vendor (Technical Safety Services, Inc. www.techsafety.com) who certifies each BSC **annually**.

The certification is conducted in accordance with NSF Standard 49 and currently accepted best practices.

3.3.3. Aerosol Management – The modified Baker SteriGard cabinet contains a separate Aerosol Management System to remove and filter aerosols from the cytometer’s interior compartments. The unit is integrated into the cabinet and uses the cabinet’s filters. This unit is used to quickly purge the sort chamber in the event of an instrument clog.

3.4. Biohazard Labels

All equipment used for storage of infectious agents must have biohazard labels specifying the agent(s) stored.

4. Facility Entry and Exit

4.1. Refer to SOP FLOW-101

5. Training

Prior to being allowed independent access to or performing work independently in the facility, all personnel will be trained by an approved lab user and must be approved by the Director of the OCS Cytometry Core.

Training will include knowledge of the Safety Manual and approved protocols, followed by observation of a certified user performing the intended procedures. Then the trainee will work under supervision of a certified lab user until the certified user gives approval and has successfully completed all training requirements outlined in Form FLOW-101F (*BSL-2 Sorting User Approval*). A copy is at the end of this manual), for the new user to be certified to enter the core or work independently in the facility.

6. Medical Requirements, Surveillance, and Responses to Exposure

6.1. Medical Requirements

Workers with a known immunodeficiency disease or who are taking immunosuppressive medications are not permitted to work in the sorting room without prior approval by the OCS Cytometry Core Director, EH&S, and Occupational Health Services. Workers with open wounds that cannot be adequately covered cannot work in the sorting room. Occupational Health Services can provide medical advice to workers who are not sure whether they fall into any of these exclusionary categories.

6.2. Medical Precautions and Surveillance

All NYULMC personnel working with patient samples are offered a Hepatitis B (HBV) vaccination at their Employee Health Screening. Personnel beginning to work with bloodborne pathogens or other potentially infectious materials can obtain an HBV immunization by contacting Occupational Health Services at 212-263-5020. Occupational Health Services can also test whether an HBV immunization is still effective.

Baseline HIV testing is required before working with biohazardous HIV-infected samples. HIV testing can be obtained at the NYULMC Occupational Health Services by contacting them at 212-263-5020. HIV testing is required subsequently in cases of accidental or suspected exposure. Personnel are also encouraged to speak to their primary care physicians about regular HIV testing.

Non-NYULMC employees are responsible for maintenance of their HBV immunizations and

for their own HIV testing.

6.3. Medical Response to Exposure

Procedures for management of exposure due to cuts are detailed in Safety Policy 135, *Bloodborne Pathogens Exposure Control Program*. These procedures apply to all NYULMC employees and as such apply to all employees working in the OCS Cytometry Core. These procedures are stated below.

All cuts and other exposures to blood or other body fluids must be reported *immediately* to Occupational Health Services, or, if not open at that time, to the Emergency Department (ED) in the Tisch building. The worker should also notify the OCS Cytometry Core Director and the Study Principal Investigator as soon as possible, after appropriate emergency care has been obtained.

Follow-up treatment for all exposures in the OCS Cytometry Core will be advised by, offered by or arranged by Occupational Health Services or the ER. Form FLOW-102F BSL-2 Spill/Accident Report should be filed with the Laboratory Manager.

7. Standard Operating Procedures (SOPs) for the OCS Cytometry Core

7.1. A dedicated set of SOPs and forms are to be followed and used by all personnel using the OCS Cytometry Core.

The currently approved SOPs and forms pertaining specifically to the virus laboratory are on the following pages of this safety manual and are listed below:

Document No.	Name
SOP FLOW-101	Standard Laboratory Practices for the OCS Cytometry Core Skirball Cell Sorting Laboratory
SOP FLOW-102	Spill Response and Reporting
SOP FLOW-103	Exposure Incidents and Reporting
SOP FLOW-104	Shipping and Receiving Infectious Substances and On-campus Transportation of Biological Samples
SOP FLOW-105	Medical and Facility Emergencies
Form FLOW-101F	BSL-2 Sorting User Approval
Form FLOW-102F	Core Facility Spill/Accident Report

Standard Operating Procedures**Title:** Standard Laboratory Practices for the OCS Cytometry Core Skirball Cell Sorting Laboratory**SOP#:** FLOW-101**Purpose:** To provide safe handling procedures and operations for all personnel working in the facility**1. Materials**

Item	Manufacturer	Catalog No.
Lab Coats	-	NYU Building Services
Safety Glasses	Jackson Safety	19706-002
Gloves	Evolution One	Small EV-2050-S Medium EV-2050-M Large EV-2050-L
200 Proof Ethyl Alcohol	-	NYU Requisition
Bleach	Clorox	Staples# CLO 02489
Kim Wipes	KimTech	34155
FACSFlow Sheath	BD	342003
Alcohol Swabs	BD	366894
Chicken Red Blood Cells Glutaraldehyde-fixed following protocol in Ref.	Sigma	R0504
Align Flow Beads Red	Molecular Probes	A16504
Align Flow Beads Yellow/Green	Molecular Probes	C16509
Align Flow Beads UV	Molecular Probes	A16502
SortCal	Sony BioTech	700002
GloGerm Beads	GloGerm	

2. Restricted Access

- 2.1. Entry into the OCS Cytometry Core Laboratory is restricted to authorized individuals who have received medical clearance from Occupational Health Services, have taken the *Intro to Biosafety* training, the *OSHA Bloodborne Pathogens* self study, and reviewed the SOPs for the OCS Cytometry Core Laboratory.
- 2.2. NYULMC's EH&S department will be granted access to conduct unannounced inspections.
- 2.3. Entry into the OCS Cytometry Core is restricted by a keyed lock and users must always be accompanied by a member of the Flow Core Staff unless otherwise authorized.
- 2.4. During sorting of potentially infectious agents, access to the laboratory will be restricted.

3. General Facility Requirements

- 3.1. Use of needles and other sharp instruments will not be used when biohazardous samples are present.
- 3.2. All cuts in the skin must be covered with a bandage.
- 3.3. No food or drinks are allowed.
- 3.4. No open-toed shoes are to be worn in the facility.
- 3.5. No jewelry (other than wedding bands) is to be worn under gloves.

- 3.6. No mouth pipetting is allowed in the facility.
- 3.7. All samples must be labeled with name, date and specimen type with a water/alcohol resistant marker.
- 3.8. Post-sort clean up should follow procedures listed in the *Decontamination* and *Exit out of the OCS Cytometry Core* sections of this SOP.

4. Reagents and Supplies

- 4.1. All samples that are transported to the OCS Cytometry Core must be contained using approved secondary containers. Refer to SOP FLOW-104
- 4.2. Unopened, non-infectious, non-toxic reagents and supplies may be stored in the front room of SK 3-9.

5. Entry to the OCS Core Laboratory and Personal Protective Equipment (PPE)

- 5.1. All outside clothing not worn under a lab coat must be left in the front room of room SK 3-9. Bags and other personal effects not to be used in the Core Laboratory should be left in SK 3-9 front room as well.
- 5.2. Wearing two pairs of gloves is advisable. They are disposed when overtly contaminated and removed when work is completed or integrity is compromised. Small, medium and large (latex or nitrile) gloves are available to the right of the entrance to the room, and should be worn at all times. They will be sprayed with 70% ethanol or isopropyl alcohol as necessary and are not to be worn outside the lab.
- 5.3. Lab coats are available on a coat rack next to entrance of the back room of SK 3-9 BSL-2 laboratory and are to be worn at all times while inside the lab. If a different size is needed, coveralls or surgical gowns can be supplied. Non-disposable lab coats are laundered on a regular basis by NYULMC Building Services.
- 5.4. The laboratory door should remain closed except when entering and exiting the lab.

6. Aerosol generating procedures

All transfers of biohazardous materials from one container to another container must take place within a BSC. Such transfers may **not** take place on the open bench.

All other procedures that could generate aerosols must also be conducted in a BSC. The following are examples of these procedures:

- Mixing of samples with a pipette;
- Using high speed mixing devices like vortexers;
- Opening of centrifuge buckets; and
- Opening a package containing an infectious pathogen.

7. Use of SY3200 Flow Cytometer and modified Baker SteriGard BSC

Considering the containment measures in place for the Sony SY3200 cell sorter (see section 7.2), the sorter's use is not considered a high-risk procedure. However, in order to ensure optimal safety, certain procedures need to be followed during setup, use and shutdown of the

instrument.

The Sony SY3200 cell sorter is operated according to the manufacturer’s manual (available at NYULMC OCS Flow Cytometry Facility Core).

7.1 Startup protocol

Machine startup steps should be performed in the following order:

Procedure	Comments
Check sheath, waste and diH2O levels.	Sheath should not be added unless waste tank is emptied. See section 7.4.1 for emptying waste. Add sheath and diH2O as necessary.
Turn on Air and Vacuum lines.	Ensure vacuum filter is dry. Replace if necessary.
Turn on BSC and Sony SY3200 cell sorter.	Machine and blower should be on for 15 minutes before use.
Run “Startup” using the SY3200 software.	Verify vacuum system is clearing waste stream
Align and setup sorter as per the manufacturer’s user manual.	

No sample should be placed on the sorter until these steps have been completed.

7.2 Clog or failure protocol

In the event of a clog, the stream may lose stability and generate an aerosol. The stream should be turned off immediately and the sample removed. Any aerosol should be contained within the sort block and filtered out by the Aerosol Management System (AMS). If the stream restarts correctly, the sort can continue. If the clog persists, the nozzle must be removed and cleaned or replaced with a secondary stand-by nozzle

If the nozzle must be removed for cleaning, it should be treated as biohazardous. After determining that the nozzle needs to be removed, the operator should activate the AMS and wait 60 seconds for any aerosol to clear from the interior compartments. The stream should be turned off, and the nozzle taken out of the machine. The nozzle should be placed into a 5mL tube, filled with 10% Contrad detergent, which is then capped. This tube is then placed into the sonicator for cleaning. After sonication, the capped tube is opened and the cleaned nozzle can be removed once back in the BSC.

Prevention of clogs by filtering all samples through a 70um cell strainer (or smaller) greatly reduces the likelihood of aerosol generation, and is therefore required. Samples should be filtered prior to delivery to operator, though strainers will be available in the sorting room to re-strain particularly problematic samples.

In the event that the vacuum line fails, an aerosol can be created where the stream enters the waste catcher. Any aerosol should be contained within the sort block and filtered out by the AMS. Turn the stream off immediately and attempt to resume house-supplied vacuum. If unsuccessful, the sorter cannot be used until Facilities has resolved the issue.

7.3 Shutdown protocol

After sorting is completed, follow the protocol below for sort shutdown:

Procedure	Comments
Remove Collection tubes from sort chamber.	Wait at least 60 seconds after stream is turned off before opening sorting chamber in order to allow aerosol evacuation.
Remove all sample and infectious material from sorter and hood.	All waste tubes should be capped and placed in waste bag. Waste bag should be sealed and sprayed before removal.
Run a tube of 10% bleach for 10 minutes and run the sample line decontamination procedure (7.3.1)	This will decontaminate all sample tubing
Wipe down all surfaces with 70% ethanol or RelyOn wipes.	Both surfaces inside and outside the hood should be decontaminated as in Section 7.4.2.
Empty waste tank if full.	Waste needs to be decontaminated before disposal. See section 7.4.1
Shutdown software, Synergy, and air and vacuum lines.	

7.3.1 Sample line decontamination procedure

In the Sony SY3200 Software ensure the flow is switched from Sheath to Rinse. Load a tube filled with 10% bleach to a volume greater than that of the sample just run. Load and run the bleach through the sample lines. Toggle sample flow on and off to ensure that the entire nozzle is disinfected. Repeat above procedure with a tube of diH2O to rinse out the bleach.

This procedure ensures that ALL tubing that is exposed to sample during regular operation is disinfected. These surfaces include the sample tubing, the pinch valve tubing and surfaces in the flow body and nozzle.

7.4 Maintenance/decontamination

7.4.1 Sony SY3200 cell sorter waste disposal

All material entering the sorter's waste tank should be considered biohazardous and must be disinfected before disposal down the sink in the cell sorting laboratory. To accomplish this, ~1L of Wescodyne disinfectant must be added to the waste tank before use. This ensures that the Wescodyne concentration will be high enough to decontaminate fully when the tank is completely filled. Wescodyne solution must be in contact with contaminated waste fluid for a minimum of 30 minutes before disposal, after which the waste fluid can be disposed of in the lab sink. The inside of the tank should be visually inspected after emptying to confirm integrity has not been compromised.

7.4.2 Surface disinfection

After sorting all surfaces inside the hood and in the Synergy's interior compartments should be disinfected with 70% ethanol.

Surfaces outside the hood that may have accidentally become contaminated should be decontaminated with 70% ethanol as well. This includes computer desk surfaces, as well as the keyboard and mouse. If the protective covers on the keyboard and mouse have become soiled, they should be disposed of and replaced before the next sort.

7.4.3 SY3200 fluidics decontamination

This procedure should be run on a monthly basis or when the machine's fluidics have become contaminated.

1. Remove sheath and diH₂O filters and replace with bypass connectors.
2. Fill sheath and diH₂O tanks with 10% Bleach solution.
3. Re-pressurize the fluidics and start stream with liquid from the rinse tank for 15 minutes.
4. Switch over to liquid from the sheath tank for 15 minutes.
5. Run 10% bleach sample line during steps 3 and 4, cycling all valves to ensure they are cleaned appropriately.
6. Empty both the sheath and waste tanks rinse with diH₂O.
7. Fill both tanks with diH₂O.
8. Run stream with diH₂O for 15 minutes from sheath tank and then 15 minutes from waste tank. Run diH₂O on the sample port during these rinses.
9. Refill diH₂O tank with diH₂O and the sheath tank with sheath fluid.
10. Replace sheath and diH₂O filters with new filters and prime system.

7.5 Aerosol containment test

In order to ensure containment of aerosols by the Aerosol Management System (AMS) and BSC, the aerosol containment testing protocol should be used at a minimum after service of the Sony SY3200 cell sorter.

The equipment used in this protocol include a fluorescence microscope, and Glo Germ™ beads, as well as an E-Lite Pump™ and an Air-O-Cell™ cassette, both supplied by EMSL Analytical, Inc. The test protocol is adapted from the sampling guide available from the manufacturer. The protocol follows:

- Prior to sampling, calibrate the pump to 15 liters per minute.
- Remove and retain tape seal covering Air-O-Cell™ inlet and outlet.
- Attach the outlet (round hole) to the supplied tubing adapter, positioning the inlet within 1 foot of the sort chamber. The sort chamber cover should be removed.
- Begin sorting Glo Germ™ beads at 20,000 events per second and simulate an aerosol causing clog by moving the waste catcher.
- Start the sampling pump, and sample for 10 minutes.
- Remove Air-O-Cell™ from tubing, and reseal with the original tape. Label sample. For a positive control, the above steps should be repeated with the aerosol containment tubing pinched closed.
- Extract coverslip from inside of cassette, lay on slide and image using the fluorescent microscope. Count Glo Germ™ beads seen.

Aerosol containment is considered maintained when less than 2 beads are present per coverslip. Ensure that the positive control is run AFTER the test sample.

7.6 Service

Field service engineers when working on the SY3200 are required to abide by this SOP.

8. Use of MoFlo XDP Flow Cytometer and Walker Class II BSC

Considering the containment measures in place for the Beckman Coulter (BCI) MoFlo XDP cell sorter (see section 8.2), the sorter’s use is not considered a high-risk procedure. However, in order to ensure optimal safety, certain procedures need to be followed during setup, use and shutdown of the instrument.

The BCI MoFlo XDP cell sorter is operated according to the manufacturer’s manual (available at NYULMC OCS Flow Cytometry Facility Core).

The startup and operating procedure consists of four stages:

1. Setup procedure to be carried out with the BSC front panel door open – supply fluid replacement and fluid waste disposal; laser alignment; drop delay calibration; side stream aiming; and cleaning with 70% ethanol solution to establish aseptic conditions inside the BSC.
2. Closing the BSC front, and equilibrating laminar airflow to establish BSL-2 cell sorting conditions.
3. Standard operation of the MoFlo XDP with the Walker BSC with front door panel closed, and airflow on.
4. Procedure in the event of a nozzle clog, or instrument malfunction and shutdown procedure.

8.1 Startup protocol

Machine startup steps should be performed in the following order:

Step	Procedure	Comments
1.	Turn on Walker BSC and BCI MoFlo cell sorter.	With the BSC front closed, including keeping the front access guard panel closed, turn BSC power switch to “Airflow On” or “Airflow + Light On.” After 1 minute with BSC airflow on, remove front access guard panel.
2.	Open front panel door of BSC and proceed with MoFlo setup.	Ensure that the BSC airflow alarm that is present briefly on startup shuts off, and that the green led lights near the power switch are illuminated, thus indicating safe airflow.

3.	Check sheath, and waste levels.	Sheath should not be added unless waste tank is emptied. See section 8.4.1 for emptying waste. Add sheath and empty waste as necessary.
4.	Turn on JunAir air compressor.	Ensure that the waste container gauge reads 20 – 25psi of vacuum, and that sheath supply tank gauge reads 30 or 60psi, depending on which nozzle is installed in the MoFlo.
5.	Verify installed nozzle size; change to correct nozzle if needed.	
6.	Turn stream on, and start Summit (MoFlo control) software.	Verify vacuum system is clearing waste stream.
7.	Align and setup sorter as per the manufacturer's user manual.	
8.	Establish proper side streams, and complete Intellisort-II setup.	
9.	Change necessary optical filters.	Rear BSC hatches must be replaced after changing optical filters, and before running biohazardous samples.
10.	Ensure BSC hatches are closed, airflow L.E.D.s indicate safe airflow and proper PPE is used.	
11.	Begin running samples/sorting.	

Steps 6, 7, 8 and 9 must be completed before a biohazardous sample is run on the instrument. These steps require open hatches on the hood and can create aerosols.

8.2 Clog or failure protocol

In the event of a clog, the stream may lose stability and generate an aerosol. The stream should be turned off immediately and the sample removed. If the stream restarts correctly, the sort can continue. If the clog persists, the nozzle must be removed and cleaned or replaced with a secondary stand-by nozzle.

If the nozzle must be removed for cleaning, it should be treated as biohazardous. After determining that the nozzle needs to be removed, the operator must wait 20 minutes before opening the BSC front panel door for any aerosol to clear from the interior compartments. The stream should be turned off, and the nozzle taken out of the machine. Within the BSC, the nozzle should be placed into a 5mL tube, filled with 10% Contrad detergent, which is then capped. This tube is then placed into the sonicator for cleaning. After sonication, the capped tube is opened within the BSC and the cleaned nozzle can be replaced.

Prevention of clogs by filtering all samples through a 70um cell strainer (or smaller) will greatly reduce the likelihood of aerosol generation, and is therefore required. Samples should be filtered prior to delivery to operator, though strainers will be available in the sorting room to re-strain particularly problematic samples.

In the event that the vacuum line, or vacuum pump fails, an aerosol can be created where the stream enters the waste catcher. Turn the stream off immediately and attempt to resume house-supplied vacuum. If unable, the sorter should not be used until Facilities has resolved the issue.

8.3 Shutdown protocol

After sorting is completed, follow the protocol below for sort shutdown:

Procedure	Comments
Remove Collection tubes from sort chamber.	Wait at least 60 seconds after stream is turned off before opening sorting chamber in order to allow aerosol to settle.
Remove all samples and infectious material from sorter and hood.	All waste tubes should be capped and placed in a red biohazard waste bag. The waste bag should be sealed and sprayed before removal.
Run a tube of 10% bleach for 10 minutes and run the sample line decontamination procedure (8.3.1)	This will decontaminate all sample tubing.
Shutdown Moflo according to manufacturer manual.	
Wipe down all surfaces with 70% ethanol.	Both surfaces inside and outside the hood should be decontaminated as in Section 8.4.2.
Empty waste tank if full.	Waste needs to be decontaminated before disposal. See section 8.4.1
Shutdown Junair Compressor.	
Turn off Walker BSC.	Replace front access guard panel with airflow on. Airflow alarm will sound, and BSC can be shutdown.

8.3.1 Sample line decontamination procedure

Load a tube filled with 10% bleach to a volume greater than that of the sample just run. Load and run the bleach through the sample lines. Increase sample pressure either by pushing the “boost” button on the MoFlo pressure console, or by manually increasing sample pressure via the adjustment knob on the pressure console. Repeat above procedure with a tube of diH₂O to rinse out the bleach. Lastly, repeat this procedure with a sample tube of 10% Contrad detergent in diH₂O.

This procedure ensures that ALL tubing that is exposed to sample during regular operation is disinfected. These surfaces include the sample tubing, the pinch valve tubing and surfaces in the flow body and nozzle.

8.4 Maintenance/decontamination

8.4.1 Beckman Coulter MoFlo XDP cell sorter waste disposal

All material entering the sorter’s waste tank should be considered biohazardous and must be disinfected before disposal down the sink in the cell sorting laboratory. To accomplish this, 300 - 400mL of Wescodyne disinfectant must be added to the waste tank before use. This ensures that the Wescodyne concentration will be high enough to decontaminate fully

when the tank is completely filled. Wescodyne solution must be in contact with contaminated waste fluid for a minimum of 30 minutes before disposal, after which the waste fluid can be disposed of in the lab sink. The inside of the tank should be visually inspected after emptying to confirm integrity has not been compromised.

8.4.2 Surface disinfection

After sorting all surfaces inside the hood and in the MoFlo's interior compartments should be disinfected with 70% ethanol.

Surfaces outside the hood that may have accidentally become contaminated should be decontaminated with 70% ethanol as well. This includes computer desk surfaces, as well as the keyboard and mouse. If the protective covers on the keyboard and mouse have become soiled, they should be disposed of and replaced before the next sort.

8.4.3 MoFlo fluidics decontamination

This procedure should be run on a monthly basis or when the machine's fluidics have become contaminated.

1. Remove sheath filter holder, empty sheath fluid from filter and holder, and fill filter holder containing filter with 10% bleach. Re-install filter in MoFlo.
2. Fill sheath tank with 10% Bleach solution, empty waste tank
3. Re-pressurize the fluidics system, start stream.
4. Run 10% bleach for 15 minutes, cycling all valves to ensure they are cleaned appropriately. Run 10% bleach on the sample port during this wash.
5. Empty both the sheath and waste tanks, rinse with diH₂O – at least 4 changes of diH₂O for sheath tank.
6. Remove sheath filter holder, rinse filter and filter holder thoroughly with diH₂O – at least 4 changes of diH₂O. Fill sheath filter and holder with diH₂O and re-install sheath filter in MoFlo.
7. Run stream with diH₂O for 30 minutes. Change diH₂O in sheath tank. Run stream with diH₂O for 30 minutes. Run diH₂O on the sample port during these rinses.
8. Empty remaining diH₂O from sheath filter and holder, and sheath and waste tanks. Re-fill sheath filter holder, and sheath tank with sheath fluid.
9. Re-pressurize the fluidics system, and run sheath to eliminate air bubbles from the system.

8.5 Aerosol containment test

In order to ensure containment of aerosols by the Moflo and BSC, the aerosol containment testing protocol should be used at a minimum after service of the Walker BSC.

The equipment used in this protocol include a fluorescence microscope, Glo Germ™ beads, as well as an E-Lite Pump™ and an Air-O-Cell™ cassette, both supplied by EMSL Analytical, Inc. The test protocol is adapted from the sampling guide available from the manufacturer. The protocol follows:

- Prior to sampling, calibrate the pump to 15 liters per minute.
- Remove and retain tape seal covering Air-O-Cell™ inlet and outlet.
- Attach the outlet (round hole) to the supplied tubing adapter, positioning the inlet within 1 foot of the sort chamber. The sort chamber cover should be removed.
- Begin sorting Glo Germ™ beads at 20,000 events per second and simulate an aerosol causing clog by covering the waste catcher.
- Start the sampling pump, and sample for 10 minutes.
- Remove Air-O-Cell™ from tubing, and reseal with the original tape. Label sample. For a positive control, the above steps should be repeated with the aerosol containment tubing pinched closed.
- Extract coverslip from inside of cassette, lay on slide and image using the fluorescent microscope. Count Glo Germ™ beads seen.

Aerosol containment is considered maintained when less than 2 beads are present per coverslip. Ensure that the positive control is run AFTER the test sample.

8.6 Service

Field service engineers when working on the MoFlo XDP are required to abide by this SOP.

9. Use of Baker SteriGard III Advance Class II Type A BSC

- 9.1. Before working in the BSC, the blowers and fluorescent light are switched on, and a biohazard bag, a spray bottle of 70% isopropyl alcohol, wipers, and pre-saturated wipes are placed in the cabinet. The blowers must be left on for 15 minutes before use.
- 9.2. All materials needed to complete the experiment are placed in the cabinet to limit the number of times hands pass through the air barrier. Equipment is not to be placed on the intake grills at the front of the cabinet, nor blocking the exhaust opening at the back of the cabinet.
- 9.3. A biohazard bag should be present in the cabinet. Absorbent material (such as a dry clean room wiper) is placed in the bottom of the biohazard bag. This bag is used for discarding solid waste (gloves, plastic waste, pipette tips). Once the bag is full, it is closed, wiped with 70% isopropyl alcohol and taken out of the cabinet to be collected into a larger covered waste container next to the cabinet.
- 9.4. Liquid waste should be put into a dedicated container inside the BSC with sufficient bleach to achieve a final concentration of 10% and allowed to react for a minimum of 30 minutes before disposal. Wipe or spray the outside of the container with 70% ethanol or isopropyl alcohol before removing it from the cabinet. The decontaminated liquids are then disposed of down the sink and flushed with large amounts of tap water.
- 9.5. Vacuum waste flasks should contain enough bleach to result in a 10% solution. They should never be filled more than 50%. An in-line vacuum filter must be present between the flask and the vacuum source.
- 9.6. Contaminated pipettes should be disposed of in the biohazard bags.

- 9.7. Anything removed from the BSC during the work session is to be decontaminated by wiping with 70% isopropyl alcohol while still in the BSC.
- 9.8. At the end of each work session, culture tubes, racks and other material to be removed from the cabinet are decontaminated by wiping with 70% isopropyl alcohol while still within the BSC.
- 9.9. The wipers used during cleaning along with the outer gloves are placed into a biohazard bag while still within the BSC. Wipe or spray the outside of the bag with 70% isopropyl alcohol. Place the bag into a larger covered biohazard waste container next to the cabinet.
- 9.10. A fresh pair of outer gloves is donned and the hood is now wiped down completely with 70% isopropyl alcohol.
- 9.11. All tissue or cell culture related materials should be disposable whenever possible. Only disposable plastic pipettes and plastic tubes are to be used in the facility.

10. CO₂ Incubators

The following is a list of safety practices and procedures for doing work involving the use of cell culture incubators.

- 10.1. Flasks and culture plates shall be carried to and from the incubator using plastic secondary containers.
- 10.2. In the event of bacterial or fungal contamination in the incubators, flasks and culture plates shall be moved to a BSC. Shelves shall be wiped down with 70% isopropyl alcohol and shelves should be sterilized in an autoclave.
- 10.3. Gloves must be worn when handling cultures.
- 10.4. Prior to maintenance, equipment must be decontaminated.

11. Centrifuge

The following is a list of safety practices and procedures for doing work involving the use of centrifuges.

- 11.1. Rotor buckets and lids shall be sprayed with 70% isopropyl alcohol and placed in the BSC prior to loading.
- 11.2. Samples shall be loaded into rotor/rotor buckets and sealed with the cap for the rotor bucket while in BSC.
- 11.3. After centrifuging, rotor/rotor buckets shall be moved to BSC to unload samples. Samples shall NOT be unloaded in the open room.
- 11.4. Centrifuge and rotor chambers shall be disinfected with 70% isopropyl alcohol soaked wipers following use.
- 11.5. Prior to maintenance, equipment must be decontaminated.

12. Decontamination

Work surfaces are to be decontaminated on completion of work, after any spill or splash, or when switching over to a new patient or product batch.

Decontaminate as follows:

- 12.1. **Bench tops and external equipment surfaces:** Work surfaces are wiped down with 70% ethanol or RelyOn Disinfectant Wipes.
- 12.2. **Water baths:** Water baths are completely emptied of water and wiped down with 70% ethanol or RelyOn Disinfectant Wipes.
- 12.3. **Biosafety cabinet work surfaces:** BSC work surfaces are sprayed with disinfectant cleaner (RelyOn Multi-purpose disinfectant cleaner or equivalent), allowing a 10 minute contact time, followed by wiping down with 70% isopropyl alcohol to remove excess disinfectant residue.
- 12.4. **Interior surfaces of equipment:** Interior surfaces of centrifuges (including centrifuge buckets), incubators and other large equipment are wiped down with 70% ethanol or RelyOn Disinfectant Wipes. Equipment is to be decontaminated prior to maintenance
- 12.5. **Liquid Waste:** Liquid biohazard waste will be decontaminated with sufficient bleach to achieve a final concentration of 10% for a minimum of 30 minutes and then emptied into the sink.
- 12.6. **Other Potentially Contaminated Waste:** All other potentially contaminated waste such as disposable lab coats and gloves are collected in red bags in containers with lids. Clothing that becomes contaminated with potentially infectious material will be decontaminated by spraying with 70% ethanol before being laundered or discarded.
- 12.7. All red bags containing contaminated wastes must be double-bagged and securely sealed with tape. All sharps containers should be locked closed. Outside surfaces of both red bags and sharps containers must be wiped down with disinfectant cleaner (RelyOn Multi-purpose disinfectant cleaner or equivalent) before transporting out of the lab.

13. Use of Chemicals

- 13.1. The same practices and training requirements will apply to the use of chemicals as in all other laboratories of NYULMC. Specifically, personnel must be current with *Chemical Hygiene* and *Hazardous Waste* training requirements. EH&S offers training on the 2nd Thursday of each month.
- 13.2. For all chemicals used in the facility, the user must give the Laboratory Manager a corresponding Safety Data Sheet (SDS). All personnel must be instructed as to their importance and their location within the facility; the Laboratory Manager will be in charge of monitoring chemical storage and use within the facility.

14. Disposal of hazardous chemicals

- 14.1. Hazardous chemicals will be collected in properly labeled containers in a designated area in the lab Arrangements for the disposal of hazardous chemical waste may be made by contacting EH&S.
- 14.2. Biohazard waste **cannot** be discarded through the Hazardous Waste Disposal Program.

- 14.3. Arrangements for the disposal of hazardous chemical waste that is also a biohazard may be made by contacting EH&S.

15. Exit out of the OCS Cytometry Core

- 15.1. All persons leaving the Core laboratory must remove PPE and wash hands before exiting.
- 15.2. Solid biohazard waste (red bags and sharps containers) should be stored in designated area, as there is no regular pickup and NYULMC Environmental Services must be notified for pickup.
- 15.3. Decontaminated liquid biohazard waste should be emptied into the sink and flushed with large amounts of tap water. (Refer to 11.5 in FLOW-101 for proper liquid decontamination practices).
- 15.3.1. Used liquid waste canisters should be disposed in red bag waste.
- 15.3.2. Secondary containers for carrying liquid waste containers are disinfected by spraying down with 70% ethanol or isopropyl alcohol, and may be autoclaved if needed.
- 15.4. Dispose gloves in a biohazard waste receptacle (red bag waste) and wash hands before exit.

Standard Operating Procedures
Title: Spill Response and Reporting
SOP#: FLOW-102
Purpose: To provide safe procedures for spill response in the facility

1. Materials

Item	Manufacturer	Catalog No.
Biohazard Bags	Lab Guard	19075388E
RelyOn disinfectant cleaner	Dupont	Fisher # 19-120-3881
Blue absorbent pads	Fisherbrand	14-206-62
Spill Kit	Spill Defense	25916

2. Spill Response

2.1. Spills will be decontaminated promptly by the responsible party.

2.2. Personnel in the immediate area will be alerted and access to the contaminated area (around the spill) will be clearly marked with the biohazard floor sign and restricted.

3. Spill Clean-Up

3.1. Don a lab coat, two pairs of gloves, and eye or face protection

3.2. For chemical spills, use the spill kit to clean up the spill. The spill kit contains absorbent packets and pads. For bio-hazardous, non-chemical hazard spills decontaminate with bleach or other disinfectant and paper towels or absorbent pads can be used.

3.3. Carefully cover the entire spill with an absorbent.

3.4. Taking care to avoid splashing pour a freshly prepared 1 in 10 dilution of bleach around the edges of the spill.

3.5. Allow a 30 minute contact time.

3.6. Pick up any glass with tongs

3.7. Use dry clean room wipers or the absorbent pads to wipe up the spill working from the edges into the center.

3.8. Disinfect the spill area by spraying thoroughly with RelyOn disinfectant/cleaner, allowing a 10 minute contact time before wiping dry.

3.9. Discard waste and any contaminated PPE in a red biohazard bag.

3.10. Wash hands.

4. Reporting

Spills or accidents will be reported to the EH&S, the Core Laboratory Manager, and the Director of the OCS Cytometry Core. Fill out the *Core Facility Spill/Accident Report* Form, (Form **FLOW-102F** – a copy is at the end of this safety manual or can be obtained from the Laboratory Manager) to document large spills or other potentially serious accidents.

Standard Operating Procedures**Title:** Exposure Incidents and Reporting**SOP#:** FLOW-103**Purpose:** To provide safe procedures for accidental exposures**1. Emergency Procedures:**

All personnel who work in the lab will be familiar with the Emergency Response Guide for New York University Medical Center Laboratories that is posted in the lab next to the entrance. This gives basic information on responding to fire alarms, chemical or biological spills or personal injury.

2. Exposure Incidents

Manage exposure incidents such as cuts with contaminated instruments, or splash to mucous membranes as follows:

2.1. For cuts with contaminated instruments:

- 2.1.1. Stop work immediately.
- 2.1.2. Remove contaminated gloves and allow the wound to bleed freely for a minute under warm running water.
- 2.1.3. Wash the wound with soap and water for at least 5 minutes and apply sterile gauze or a bandage, if necessary.
- 2.1.4. Remove protective lab clothing and proceed immediately to the appropriate location for treatment and counseling.

2.2. For splashes to mucosal membranes:

- 2.2.1. Stop work immediately and proceed immediately to the eye wash station.
- 2.2.2. Rinse tissue surface with copious amounts of water. Eyes should be irrigated for at least 15 minutes.
- 2.2.3. Remove protective lab clothing and proceed immediately to the appropriate location for treatment and counseling.

Appropriate Locations for Treatment and Counseling

Department	Phone Number	Location	Hours of Operation
Occupational Health Services	212-263-5020	1 Park Avenue on the 3 rd Floor	M-F 8:00AM-5:00PM
NYULMC Emergency Department	212-263-5550	530 First Avenue, HCC 102	Open 24 hours 7 days/week

Note: If a laboratory worker has a parenteral (e.g. percutaneous injury or contact with non-intact skin) or mucous membrane exposure to blood, body fluid, or viral-culture material, the source material will be identified and, if possible, tested for the presence of virus. **In general, materials handled in the OCS Cytometry Core should be considered contaminated**

unless known otherwise.

For work involving HIV-infected or potentially infected products, the worker must be escorted directly to the emergency room for **immediate** evaluation and counseling with regard to the risk of infection. **Post-exposure prophylaxis (PEP) should be offered according to the latest guidelines, and if deemed necessary, should begin as soon as possible, typically within hours of exposure.** Administration of PEP should not be delayed for HIV test results. As of August 2008, the CDC recommendation is as follows:

“Use of PEP with antiretroviral medications, initiated as soon as possible after exposure and continuing for 28 days, has been associated with a decreased risk for infection following percutaneous exposure in health-care settings (22)...Because of the potential toxicities of antiretroviral drugs, PEP is recommended unequivocally only for exposures to sources known to be HIV-infected. The decision to use PEP following unknown-source exposures is to be made on a case-by-case basis, considering the information available about the type of exposure, known risk characteristics of the source, and prevalence in the setting concerned.” [MMWR Aug 1, 2008 / 57(RR06); 1-19].

The worker will be evaluated serologically for HIV and advised to report and seek medical evaluation of any acute febrile illness that occurs within 12 weeks after the exposure. Such an illness – particularly one characterized by fever, rash, or lymphadenopathy – may indicate recent HIV infection. If the initial (at time of exposure) HIV test is negative, the worker should be retested 6 weeks after the exposure and periodically thereafter (i.e., at 12 weeks and 6, 9 and 12 months after exposure). During this follow-up period exposed workers should be counseled to follow Public Health Service recommendations for preventing transmission of HIV.

NOTE: Please note that exposure to other bloodborne pathogens or other potentially infectious materials is discussed in detail in NYULMC’s *OSHA Bloodborne Pathogens* self study.

3. Reporting

Exposure incidents must be reported immediately either in person or by phone to an OCS Cytometry Core Manager, the Director of the OCS Cytometry Core, and Occupational Health Services. Use *Core Facility Spill/Accident Report* Form, (Form **FLOW-102F** – a copy is at the end of this safety manual or can be obtained from the Laboratory Manager) to document the incident. Exposure incidents involving recombinant or synthetic DNA will be reported to the Institutional Biosafety Committee (IBC) after the above steps have been satisfied.

Standard Operating Procedures**Title:** Shipping and Receiving Infectious Substances and On-campus Transportation of Biological Samples**SOP#:** FLOW-104**Purpose:** To ensure that shipping and receiving/transportation of specimens and cultures which harbor or are suspected of harboring pathogens is performed in a controlled and dedicated manner.**1. Training Requirements**

Personnel who want to ship or receive infectious substances **must** be current with training requirements.

1.1. EH&S provides the self-study course: *Shipping Hazardous Materials*, which is available at: [iDevelop](#)
iDevelop can be accessed via atNYULMC.org

1.1 A training certificate is issued and maintained in the EH&S Department upon successful completion of both post tests mentioned in 1.1; the certification is valid for **two** years.

2. On-campus Transportation of Biological Samples**2.1. Materials**

Item	Manufacturer	Catalog No.
Biohazard Bags	Lab Guard	19075388E
Hard container (cooler)	Igloo	7362
Blue absorbent pads	Fisherbrand	14-206-62

2.2. General Notes

2.2.1. All samples and containers must have biohazard labels.

2.2.2. Avoid crowded areas whenever possible.

2.2.3. The container should be carried directly to the intended laboratory - do not take the container to offices, cafeterias or other public or inappropriate locations.

2.2.4. The package should be carefully inspected for signs of leakage or other contamination and, if necessary, decontaminated before opening.

2.3. Packaging Instructions

2.3.1. Label samples. Label information must include the identity of the biological material or agent, the universal biohazard symbol and the sending and receiving laboratory identification (*e.g., Principal Investigator name and room number*).

2.3.2. Place sample in a primary container which is sealed and leak proof.

2.3.3. Place the primary container in a secondary hard case container which is easy to decontaminate and capable of being securely closed.

2.3.4. Liquid samples should be surrounded by enough absorbent pads in the secondary container to contain any liquids and absorb any shock during transport.

Standard Operating Procedures**Title:** Medical and Facility Emergencies**SOP#:** FLOW-105**Purpose:** To provide safe procedures for handling medical and facility emergencies**1. Materials**

Item	Manufacturer	Catalog No.
Emergency flashlight	-	-
First Aid Kit	PhysiciansCare	Staples #503995

2. Medical Emergencies

- 1.1. In case of a medical emergency, call the Medical Center's emergency number: 33-911.
- 1.2. If the individual is conscious and can be moved, remove him/her immediately out of the laboratories.
- 1.3. If the individual is unconscious and it will cause no further harm, the person will be immediately removed out of the laboratories and emergency personnel will be called to perform first aid.
- 1.4. If the victim cannot be moved, instruct the emergency responders of hazards and protective measures necessary in the facility.
- 1.5. Stay with the victim until emergency medical personnel arrive and take over.

2. Electrical Failures

- 2.1. In case of a power outage the operator must use his/her own best judgment to assess the situation and act accordingly.
- 2.2. In case of an electrical failure, call NYULMC's main number for Facilities: (212) 263 5275.
- 2.3. The building's emergency power generator should mitigate any loss of power to the essential containment equipment, allowing for proper shutdown and containment of biohazards.
- 2.4. If the blower fan of a BSC stops working any operator working in the BSC is required to cease all work immediately. If possible secure any infectious material. Exit from the laboratory following the exit procedures listed in SOP FLOW-101 for removal of protective gear.
 - 2.4.1. The blower must be on for at least thirty minutes before work can resume.
- 2.5. In case of a blackout, all operators are to evacuate the facility. A rechargeable flashlight will be available for emergency use if needed.
- 2.6. Exit doors are identified with glow-in-the-dark exit signs which will allow the operator to find the exit door. Exit procedures listed in **SOP FLOW-101** will be followed.
- 2.7. A sign should be posted on the entrance door with a notice advising persons not to enter the facility.

3. Fire Emergency

- 3.1. In the event of a fire the laboratory worker must take the following steps:

- 3.1.1. If the infectious material is stored as per lab requirements, the worker removes the PPE and exits the lab quickly as per exit procedures detailed in **SOP FLOW-101**, as required when he/she leaves.
- 3.1.2. If research involving the infectious material is in progress, the worker will determine if the agent can quickly be secured or whether it is quicker to destroy the material prior to leaving the lab as outlined in the section on **SOP FLOW-101 Decontamination – Liquid Waste**.
- 3.2. After evacuating the facility on account of fire, all workers will remain at a safe distance to offer directions to the facility and any information EH&S and/or Fire Department personnel may request. When they or Fire Department personnel arrive on the scene, all workers will follow their instructions.

FORM FLOW-101F. BSL-2 Sorting User Approval

Name: _____ Date: _____

Date of birth: _____

Principal Investigator: _____ Department: _____

Title: _____ Work phone: _____

Home phone: _____ E-mail: _____

_____ I have completed, within the past year, the NYULMC training on bloodborne pathogens.

_____ I have read (and received a copy of) the OCS Cytometry Core's SOPs and am familiar with:

- Safe working practices, which all persons in the facility are expected to follow
- Appropriate responses for spills in the laboratory, both within and outside of BSCs
- Decontamination procedures
- Procedures for medical, electrical and fire emergencies
- Being familiar with the Biosafety Microbiological and Biomedical Laboratories (BMBL) manual edited by the Center for Diseases Control (CDC), especially parts concerning BSL2 work and infectious organisms being used.

_____ In compliance with the OSHA Bloodborne Pathogens Standard, NYULMC has an HBV vaccination program. I understand that under this program, any worker who is at risk from HBV from occupational exposure to human blood, blood products or body fluids, or HBV contaminated materials may receive an HBV vaccination free of charge. The HBV vaccination program is administered by Occupational Health Services.

_____ I understand that the Occupational Health Services and the ER are prepared to administer medications to reduce the risk of HIV infection following a body stick or mucous membrane exposure to HIV and that it is my responsibility to report immediately to be evaluated for such treatment in the event of a possible exposure.

User signature: _____ Date: _____

OCS Core Laboratory signature: _____ Date: _____

Form FLOW - 102F. Core Facility Spill/Accident Report

Reporting Objective:

In the process of investigating and reporting incidents the facility can determine the cause and provide recommendations for future prevention and correction of the events that lead to the accident/spill. This document is based on OSHA CLP 02-00-135-Recordkeeping Policies and Procedures Manual (effective-12/30/2004).

If additional space is needed to complete any question for a section, please attach extra page indicating which section is being continued.

1. Completed by (Name, Job Title):

2. Name/Job Title/Name of Principal Investigator: _____

3. Date/Time of Incident:

4. Infectious agent/hazardous substance involved: _____

5. Where did incident happen (which area of the Core facility)? _____

6. Describe circumstances that lead to incident (work being done at that time, location of spill, equipment involved):

7. Other persons in Core lab at time of incident (where were they; did they contribute to the incident?)

8. Duration of safety breach (time to containment): _____

9. What, if any, measures were taken to contain the safety problem?

a. Evacuation of facility ____ Yes ____ No

b. Who de-contaminated the spill (person or persons)? _____

10. Who was notified of the incident? When were they notified?

11. List any injuries as a result of this incident:
